REVIEW





Advancements in enzyme engineering for industrial biotechnology

Durgapada Sarkhel

Department of Biotechnology, Utkal University, Bhubaneshwar, Odisha

ABSTRACT

Enzyme engineering has become crucial to advancing industrial biotechnology, offering solutions across biofuel production, food processing, and environmental management. Through methods like directed evolution, rational design, and hybrid approaches, engineered enzymes demonstrate enhanced stability, specificity, and catalytic efficiency under the rigorous conditions of industrial use. These improvements enable more efficient biofuel production, high-quality food products, and effective pollutant degradation, highlighting enzyme engineering's impact on both economic and environmental sustainability. However, technical limitations, such as enzyme inactivation, high production costs, and scalability issues pose significant challenges to widespread industrial adoption. Additionally, regulatory hurdles and environmental concerns regarding genetically modified enzymes complicate commercialization efforts. Emerging technologies such as CRISPR and artificial intelligence (AI) are transforming enzyme engineering, enabling precise genetic modifications and predictive enzyme modelling, which streamline development and reduce reliance on costly experimental approaches. The growing focus on sustainable applications has led to enzymes designed for biodegradable plastic degradation and eco-friendly waste treatment, aligning with global sustainability objectives. Interdisciplinary collaboration among biotechnologists, chemists, and data scientists is driving these innovations, addressing current challenges, and expanding enzyme engineering's potential. With continued advancements, enzyme engineering is positioned to revolutionize industrial biotechnology, fostering sustainable and cost-effective solutions across various sectors. This version enhances clarity by emphasizing key points in a structured, concise manner and reinforces the potential of enzyme engineering to drive sustainable industrial practices.

Introduction

Industrial biotechnology is revolutionizing multiple sectors, from chemicals and pharmaceuticals to food and environmental management, by leveraging biological processes to create sustainable and efficient production methods. At the heart of this revolution are enzymes, which act as natural biological catalysts, promoting reactions under mild conditions that are often unattainable with traditional chemical catalysts. Key enzymes such as lipases, cellulases, and proteases are integral to processes like biofuel production, food processing, and wastewater treatment, offering significant benefits in terms of energy savings, reduced chemical waste, and eco-friendly operations [1].

However, naturally occurring enzymes are not always well-suited for industrial environments, where conditions can vary from extreme pH and temperature ranges to exposure to solvents and other harsh reagents. Enzyme engineering has thus become critical, allowing scientists to modify enzymes to enhance their stability, specificity, and activity under these demanding conditions. Tailoring enzymes through engineering not only improves their functionality but also reduces operational costs and waste, aligning with sustainability goals. Current techniques in enzyme engineering, including directed evolution, rational design, and hybrid methods, are tailored to improve specific properties that make enzymes more compatible with industrial settings, bridging the gap between

KEYWORDS

Enzyme engineering; Industrial biotechnology; Sustainable biocatalysis; Rational enzyme design; Biofuel production

ARTICLE HISTORY

Received 26 April 2024; Revised 21 May 2024; Accepted 29 June 2024

biological and industrial efficiency [2]. Directed evolution and rational design are two prominent methods used in enzyme engineering. Directed evolution replicates natural selection by creating and testing numerous enzyme variants to identify those with desirable traits, such as heightened thermal stability or enhanced catalytic rates. In contrast, rational design involves modifying specific residues within an enzyme based on its structural information, allowing for targeted improvements in properties like substrate specificity. Each approach has shown considerable success across various applications. For instance, enzymes engineered through directed evolution have enhanced biofuel yields by tolerating high-temperature fermentation processes. In contrast, rationally designed enzymes are being used in pharmaceutical synthesis to achieve precision in complex reactions [3].

Despite these advancements, challenges remain. Enzymes may still denature or lose activity under certain industrial conditions, and scaling up enzyme production can be costly and technically demanding. Moreover, directed evolution is labour-intensive and requires high-throughput screening, while rational design demands detailed structural knowledge, which is not always available. These limitations underscore the need for more refined engineering techniques and cost-effective production methods [4].

^{*}Correspondence: Mr. Durgapada Sarkhel, Department of Biotechnology, Utkal University, Bhubaneshwar, Odisha, e-mail: durgapadasarkhel98@gmail.com © 2024 The Author(s). Published by Reseapro Journals. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

This review discusses recent advancements in enzyme engineering techniques and their applications in industrial biotechnology, focusing on strategies to overcome current limitations. By exploring these developments, the review highlights the potential of engineered enzymes to drive sustainable and economically viable industrial practices.

Methods in Enzyme Engineering

Directed evolution

Directed evolution is a widely used enzyme engineering technique that imitates natural selection to improve enzyme properties like stability, substrate specificity, and catalytic efficiency. This method involves generating a library of enzyme variants through rounds of random mutagenesis, followed by screening or selection to identify variants with desired traits. Over successive iterations, enzymes can be fine-tuned to meet specific industrial demands [4,5]. Directed evolution has been transformative in applications requiring enzymes with enhanced thermal stability, such as in biofuel production. Thermostable cellulases, for example, have been developed to withstand high temperatures during biomass hydrolysis, significantly improving biofuel production efficiency. Additionally, proteases engineered for detergent formulations through directed evolution are capable of retaining activity in high-alkaline environments, a necessity for laundry detergents that require stable enzyme function in aggressive washing conditions. These examples demonstrate how directed evolution adapts enzymes to challenging industrial settings, enhancing both functionality and operational durability [6].

Several case studies highlight the success of directed evolution in optimizing enzymes for industrial use. In the biofuel industry, cellulase enzymes have been engineered to break down lignocellulosic biomass more effectively by increasing their thermal stability and catalytic activity. These optimized cellulases enable more efficient conversion of plant material into bioethanol, reducing production costs and energy requirements [4-6]. Another case study in the detergent industry involves alkaline proteases, which have been modified through directed evolution to improve performance in various washing conditions, including low-temperature and high-alkaline environments. These enzymes not only enhance stain removal but also reduce the need for synthetic chemicals, contributing to more environmentally friendly detergent formulations [7].

Rational design and computational approaches

Rational design utilizes detailed knowledge of enzyme structure and function to make targeted modifications that enhance performance in specific industrial applications. Unlike random mutagenesis in directed evolution, rational design focuses on modifying amino acids at critical sites to optimize enzyme properties like substrate specificity, stability, and activity. For instance, lipases used in the pharmaceutical industry have been engineered through rational design to achieve high enantioselectivity, allowing them to catalyse the production of chiral drug intermediates with increased precision. By understanding the enzyme's active site structure, researchers can make precise adjustments that improve its function in these highly specialized reactions [8].

Computational tools have become essential in rational design by facilitating enzyme modelling, mutation prediction, and structural analysis. Software like Rosetta and Schrödinger's Maestro enables researchers to simulate enzyme-substrate interactions, predict the effects of amino acid substitutions, and assess enzyme stability under different conditions [4,6,7]. These tools have been instrumental in streamlining the rational design process, as they allow researchers to predict which mutations are likely to produce desired traits without extensive laboratory testing. For example, Rosetta's modelling capabilities have been used to predict stabilizing mutations in enzymes destined for high-temperature industrial applications, significantly reducing the time and cost associated with experimental trials. Machine learning algorithms are also increasingly applied to large datasets from mutagenesis studies, helping to predict mutation outcomes that enhance enzyme performance [9].

Rational design has been particularly impactful in tailoring enzymes for industry-specific needs. In the food industry, glucose oxidase has been modified to improve its stability in acidic environments, making it highly effective in food preservation and flavour enhancement. Similarly, in the pharmaceutical sector, transaminases have been engineered through rational design to improve their stereoselectivity, which is crucial for the synthesis of enantiomerically pure drugs. These enzymes are now used in the production of active pharmaceutical ingredients with increased precision and lower by-product formation, illustrating the benefits of rational design in highly regulated industries [10].

Hybrid approaches

Hybrid methods combine the advantages of directed evolution and rational design, creating a powerful approach to enzyme engineering. These methods often start with rational design to introduce foundational structural changes, followed by directed evolution to refine and enhance enzyme traits [9,10]. For instance, nitrilases used in chemical synthesis have been engineered using a hybrid approach: rational design was employed to optimize substrate binding, while subsequent rounds of directed evolution improved catalytic efficiency and stability. By leveraging both techniques, hybrid approaches enable the development of enzymes that are finely tuned for complex industrial applications, where single-method strategies might fall short [11] (Table 1).

Enzyme Engineering Approaches	Steps and Techniques	Description
	Error-Prone PCR	Introduces random mutations to generate a variety of enzyme variants.
	High Throughput Screening	Uses fluorescence-activated cell sorting to quickly analyze and
Directed Evolution	(FACS)	select desired enzyme variants.
	Analysis	Evaluates the enzyme variants to identify those with improved characteristics.

17



	Data Sets Molecular Predictors	Collects data on enzyme structures and functions for modeling purposes. Uses molecular predictors to understand enzyme behavior and potential modifications
Rational Design	Machine Learning Algorithms Hot Spot Prediction	Applies machine learning to predict outcomes of enzyme mutations. Identifies key residues in the enzyme structure to target for modification.
	Site-Directed Mutagenesis	Precisely alters specific amino acids in the enzyme to improve function or stability.
Semi-Rational Design	Sequence Analysis	Examines the sequence of enzymes to find regions for potential improvement.
	Homology Modeling	Uses similar enzyme structures to build models for predicting mutations and their effects.
	Structure Validation	Verifies the accuracy of the homology models before proceeding with mutations.
	Random and Site-Directed Mutagenesis	Combines random and specific mutations to create enzyme variants with desirable traits.
	Double Network Gel	Uses a gel matrix to provide a stable environment for enzyme immobilization.
Enzyme Immobilization	Addition of Enzyme	Introduces enzymes into the gel matrix to restrict their movement and enhance stability.
	Immobilized Enzyme	Immobilizes enzymes within the matrix to improve their operational stability and reusability.

Recent success

Recent advancements in hybrid enzyme engineering have opened new possibilities in challenging industrial sectors. One notable example is the development of enzymes for plastic degradation. Using a hybrid approach, researchers have engineered cutinases with enhanced activity and stability against polyethene terephthalate (PET), facilitating the breakdown of plastic waste a critical step toward reducing environmental pollution [11]. Another success involves cytochrome P450 enzymes engineered for bioactive compound synthesis in the pharmaceutical industry. By combining rational modifications with directed evolution, these enzymes have achieved higher yield and selectivity, enhancing their value in drug development and synthesis processes. These successes underscore the potential of hybrid methods in advancing enzyme engineering for sustainable and economically viable industrial applications [12].

Engineering Enzymes for Industrial Applications

Biofuel production

Enzymes are essential in biofuel production, particularly in the breakdown of lignocellulosic biomass for bioethanol and biodiesel synthesis. Cellulases and hemicelluloses break down complex plant structures into fermentable sugars, enabling efficient bioethanol production. Lipases, on the other hand, catalyse transesterification reactions necessary for biodiesel synthesis. However, the industrial conditions in biofuel production, including high temperatures and exposure to organic solvents, often lead to enzyme inactivation. These harsh conditions demand enzyme variants with enhanced thermal and solvent stability, which are not typically found in natural enzymes [13].

A significant challenge in biofuel production is maintaining enzyme efficiency under high-temperature conditions typical of industrial biomass reactors. Enzymes such as cellulases tend to lose catalytic activity at these elevated temperatures, leading to inefficiencies and increased costs due to enzyme replenishment. Directed evolution has been used to develop thermostable cellulase variants capable of sustained activity in high-temperature environments, which enhances the breakdown of biomass into fermentable sugars and improves overall bioethanol yield [10-12]. Similarly, for biodiesel production, lipases have been engineered using rational design to maintain activity in organic solvent environments. This modification enables lipases to function effectively during transesterification reactions, significantly improving biodiesel synthesis efficiency. These advances have not only bolstered enzyme durability but also reduced biofuel production costs, making biofuel a more viable alternative to fossil fuels [14] (Table 2).

 Table 2. Current Situation of Enzyme Engineering in biofuel production.

Aspect	Current Scenario	
Target	Cellulases and Hemicelluloses: Used for breaking down lignocellulosic biomass into fermentable sugars for	
Enzymes	bioethanol production.	
	Lipases: Utilized in biodiesel production through transesterification of fats and oils.	

18

Engineering Techniques	Directed Evolution: Random mutation and selection cycles enhance enzyme traits like thermal stability an catalytic efficiency.		
	Rational Design: Structural analysis and computational tools allow targeted modifications to specific residues.		
Technological Advancements	Use of CRISPR for precise gene editing to improve enzyme traits. AI-driven models such as AlphaFold assist in predicting enzyme structure and function, making design more efficient and cost-effective.		
Industrial Applications	Enhanced cellulases and hemicelluloses have increased bioethanol yields by enabling a more efficient breakdown of plant biomass. Engineered lipases in biodiesel production are more stable in organic solvents, improving reaction efficiency.		
Sustainability Focus	Engineered enzymes reduce reliance on harsh chemicals and lower energy consumption in biofuel production processes. Enzyme modifications aim to achieve higher stability, reducing enzyme replacement frequency and associated costs.		
Challenges	Stability: Maintaining enzyme activity at high temperatures and in varying pH levels is still challenging. Cost: High costs for enzyme production, purification, and scaling remain a barrier to widespread adoption.		
Future Directions	Emphasis on green chemistry and sustainable biofuel production. Interdisciplinary collaboration among biotechnologists, chemists, and data scientists is expected to drive further advancements in enzyme engineering for biofuels.		

Food and beverage processing

Enzymes play a critical role in the food and beverage industry, where they contribute to texture, flavour, and product stability (Table 3). In dairy processing, proteases are engineered to accelerate cheese ripening, enhancing flavour profiles through targeted protein breakdown. Lipases, adapted for specific pH and temperature ranges, aid in fat modification processes, which are essential for producing consistent flavours in cheese, yoghurt, and other dairy products [15]. Additionally, in the baking industry, amylases engineered for high-temperature stability improve the dough's fermentation process and extend bread freshness by breaking down starches into fermentable sugars. These enzyme modifications allow for better performance under baking conditions, promoting product uniformity and extended shelf life [16].

Engineered proteases and lipases have seen particular success in the food industry due to their tailored stability and activity profiles. Acid-stable proteases, for example, are designed for dairy applications, where they remain active under low pH conditions in cheese processing. These proteases accelerate flavour development without compromising product quality or safety. In baked goods, thermostable lipases facilitate the modification of fats, creating desirable textures and improving shelf life. By tailoring these enzymes to specific industrial needs, enzyme engineering has enabled the food industry to achieve consistent product quality and efficiency in production processes [17].

Table 3. Enzyme Engineering methods used in food industry.

Enzyme Engineering Approach	Techniques Used	Applications in Food Industry	Benefits
Directed Evolution	Error-Prone PCR	Development of enzymes for improved baking (e.g., xylanases, lipases)	Enhanced enzyme stability and efficiency
	High-throughput screening (e.g., FACS)	Creation of new enzymes for dairy processing to enhance flavour	Optimized enzyme activity in various pH and temperature conditions
	DNA Shuffling	Fermentation enhancements in brewing yeast	Increased production efficiency in baking and brewing
Rational Design	Molecular Modelling	Production of lactase enzymes with improved stability for lactose-free dairy	Targeted modification of enzymes for specific reactions
	Machine Learning Algorithms	Engineering proteases for specific meat tenderization	Consistent performance under industrial conditions

19



	Site-Directed Mutagenesis	Development of amylases with improved resistance in baking	Cost-effective enzyme usage for specific applications
Semi-Rational Design	Homology Modelling	Production of pectinases and cellulases for fruit juice clarification	Increased enzyme stability and longer shelf-life
	Random and Site- Directed Mutagenesis	Enhancing lipase stability in oil processing	Tailored enzymes for unique flavour profiles
		Modified enzymes for cheese ageing and flavour development	Enhanced clarity and quality in juice production
Enzyme Immobilization	Double Network Gels	Reusable lactase enzymes in lactose- free milk production	Reduced production costs due to enzyme reusability
	Enzyme Entrapment in Gels or Beads	Immobilized glucose isomerase for continuous fructose syrup production	Consistent quality in continuous processing
	Cross-Linking	Lipases for fat modification in oils	Enhanced stability and control over enzymatic reactions
Protein Engineering for Specific Functions	Surface Engineering	Engineering enzymes to withstand high temperatures in pasteurization	Enzymes capable of functioning under extreme processing conditions
	Active Site Modification	Development of lipases and proteases with specific substrate affinity for dairy and bakery applications	Higher product yield and quality Reduced need for artificial additive

Environmental bioremediation

Enzyme engineering is crucial in environmental bioremediation , where enzymes are used to degrade pollutants and treat industrial waste. Synthetic pollutants, such as plastics, pesticides, and industrial dyes, are highly resistant to natural degradation processes. Engineered enzymes, particularly laccases and peroxidases, are designed to withstand extreme pH and temperature conditions, making them suitable for breaking down these persistent contaminants [10,12]. For instance, laccases have been modified to function in alkaline environments and are effective in degrading azo dyes commonly found in textile industry wastewater. The enhanced stability and catalytic efficiency of these engineered enzymes allow them to reduce pollutant levels significantly, improving wastewater quality before discharge [18].

Several successful applications of engineered enzymes in bioremediation highlight their environmental impact. A notable case is the use of laccases for dye degradation in wastewater treatment. Engineered laccases that operate in alkaline conditions have been shown to break down synthetic dyes by cleaving azo bonds, reducing water toxicity by up to 90% and facilitating safer disposal [11-15]. Another impactful case is the application of engineered cutinases in plastic degradation. These enzymes have been modified to hydrolyze polyethene terephthalate (PET), a major component of plastic waste. Engineered cutinases can depolymerize PET under industrial conditions, providing a potential solution for recycling plastic waste and reducing environmental pollution. Such examples underscore the value of enzyme engineering in addressing pressing environmental issues [19].

Limitations in Enzyme Engineering Technical constraints

Despite significant advances in enzyme engineering, technical challenges continue to limit the scalability and functionality of engineered enzymes in industrial applications. One major issue is enzyme inactivation, which frequently occurs under the high-temperature and extreme pH conditions typical in sectors like biofuel production. For example, cellulases used in biomass conversion are prone to denaturation at elevated temperatures, leading to diminished catalytic efficiency and higher operational costs due to the need for frequent enzyme replenishment [17,20]. Another technical limitation lies in protein expression. Engineered enzymes often require specific post-translational modifications for activity, which common expression systems, such as E. coli, cannot provide. While alternative hosts like yeast and mammalian cells offer some solutions, they can be costly and present issues with yield and scalability. Large-scale production also poses substantial technical obstacles. Enzymes must be produced in vast quantities while maintaining purity and activity, but current bioprocessing systems often encounter contamination risks and loss of activity during fermentation and purification stages, adding complexity to industrial-scale applications [20].

Cost considerations

The economic feasibility of enzyme engineering remains a critical hurdle, particularly due to the high costs associated with enzyme production and purification. For instance, purification steps alone can account for up to 50% of the total production cost, as specialized equipment and protocols are necessary to

isolate active enzymes from by-products and contaminants [21,22]. Additionally, quality control measures, required to meet industry standards, contribute significantly to the overall cost, making it challenging for small and medium enterprises to adopt engineered enzymes for their operations. Recent efforts to reduce costs, such as the development of cost-effective expression systems and downstream processing innovations, show promise but have yet to fully address these economic barriers. Consequently, cost remains a primary limitation in the commercialization of engineered enzymes, often restricting their use to high-value applications [21].

Regulatory and environmental factors

Regulatory challenges add another layer of complexity to enzyme engineering, especially when using genetically modified organisms (GMOs) as hosts for enzyme production. Regulatory bodies like the U.S. Food and Drug Administration (FDA) and the European Food Safety Authority (EFSA) enforce stringent guidelines to ensure the safety of GMOs, with regulations often varying widely between regions. For example, the European Union imposes strict regulations on GMOs, which can delay or even prevent the approval of genetically engineered enzymes for food and environmental applications [22]. In addition to regulatory barriers, environmental concerns about the long-term effects of releasing engineered enzymes into ecosystems must be carefully considered. For instance, the release of enzymes engineered for waste degradation poses potential risks to natural microbial communities and could lead to unintended ecological interactions. Some studies suggest that engineered enzymes may disrupt native microbial populations or contribute to horizontal gene transfer, raising concerns about ecosystem balance. Environmental assessments and containment strategies are increasingly required to mitigate these potential risks, but they add both time and cost to the development process [23].

Future Perspectives in Enzyme Engineering

Emerging technologies

The future of enzyme engineering is being revolutionized by technologies like CRISPR and artificial intelligence (AI), which offer unprecedented precision in enzyme modification. CRISPR-Cas systems enable targeted gene editing, allowing researchers to alter specific regions within enzyme-coding genes to enhance traits such as substrate affinity, stability, and catalytic efficiency. A notable example includes the use of CRISPR to modify cellulases for higher activity and thermal stability, enhancing their efficiency in biofuel production processes [23]. AI, particularly tools like AlphaFold for protein structure prediction, has transformed enzyme engineering by allowing researchers to predict enzyme folding and function based on genetic sequences. AI-driven modelling has enabled rapid identification of beneficial mutations, improving enzyme activity and reducing the costly trial-and-error process of traditional engineering methods. Together, CRISPR and AI represent powerful tools that streamline enzyme design, accelerating advancements in industrial applications such as pharmaceuticals and sustainable chemicals [24].

Sustainable and green enzyme engineering

There is a growing emphasis on developing enzymes that

support environmentally friendly and sustainable industrial practices. Engineered enzymes are increasingly employed in processes that reduce reliance on harsh chemicals, lower energy consumption, and produce fewer by-products. For example, engineered enzymes for biofuel production can reduce carbon emissions by up to 70% compared to fossil fuels, highlighting their potential in renewable energy [21,23]. In waste management, enzymes designed for plastic degradation, like engineered PETase variants, enable the breakdown of polyethene terephthalate (PET) plastics under mild conditions, offering a solution to plastic pollution. These enzymes provide a sustainable approach to recycling plastics without relying on energy-intensive processes, making them crucial for green chemistry initiatives. By reducing the environmental footprint of industrial processes, sustainable enzyme engineering is aligning with global goals for reducing pollution and conserving natural resources [25].

Interdisciplinary collaboration

The advancement of enzyme engineering heavily relies on interdisciplinary collaboration across fields such as biotechnology, chemistry, data science, and bioinformatics. Projects like the National Institutes of Health's Protein Structure Initiative exemplify successful interdisciplinary work, bringing together structural biologists, chemists, and data scientists to design and predict enzyme behaviours for medical and industrial applications. In a recent collaboration, biotechnologists and computational scientists engineered nitrilases for pharmaceutical synthesis by combining high-throughput screening with AI-driven structural analysis, significantly enhancing the enzyme's efficiency [24,25]. Such collaborations allow each discipline to contribute specialized knowledge, enabling more rapid advancements and practical solutions to complex engineering challenges. As enzyme engineering increasingly integrates with computational and data science, its scope and impact are expected to expand, paving the way for applications in diverse sectors, from healthcare to environmental science [1].

Conclusion

Enzyme engineering has made remarkable strides through advancements in techniques such as directed evolution, rational design, and hybrid approaches, which have enhanced enzyme stability, specificity, and catalytic efficiency. These optimized enzymes have driven progress in fields like biofuel production, food processing, and environmental bioremediation, reducing operational costs and increasing process efficiency across diverse industrial applications. One of the most transformative impacts of enzyme engineering lies in its potential to support sustainable practices. By facilitating environmentally friendly processes that reduce chemical waste and energy consumption, engineered enzymes contribute to global sustainability efforts. For instance, enzymes like PETase, designed for plastic degradation, offer innovative solutions to plastic pollution, demonstrating the potential of enzyme engineering to advance green chemistry and address pressing environmental challenges. As the field continues to evolve, overcoming technical and regulatory challenges will be essential to realizing the full potential of engineered enzymes across sectors. Emerging technologies such as AI and CRISPR are expected to further

drive innovation, enhancing enzyme efficiency and regulatory compliance. Continued interdisciplinary collaboration will also be key, solidifying enzyme engineering as a cornerstone of future industrial and environmental biotechnology.

Disclosure statement

No potential conflict of interest was reported by the authors.

References

- 1. Jordan Chapman AI, Dinu C. Industrial applications of enzymes: recent advances, techniques, and outlooks. Catalysts. 2018;8(6):238. https://doi.org/10.3390/catal8060238
- Victorino da Silva Amatto I, Gonsales da Rosa-Garzon N, Antonio de Oliveira Simoes F, Santiago F, Pereira da Silva Leite N, Raspante Martins J, et al. Enzyme engineering and its industrial applications. Biotechnol Appl Biochem. 2022;69(2):389-409. https://doi.org/10.1002/bab.2117
- Porter JL, Rusli RA, Ollis DL. Directed evolution of enzymes for industrial biocatalysis. Chem Bio Chem. 2016;17(3):197-203. https://doi.org/10.1002/cbic.201500280
- Dinmukhamed T, Huang Z, Liu Y, Lv X, Li J, Du G, et al. Current advances in design and engineering strategies of industrial enzymes. Syst Microbiol Biomanufacturing. 2021;1:15-23. https://doi.org/10.1007/s43393-020-00005-9
- Martínez R, Schwaneberg U. A roadmap to directed enzyme evolution and screening systems for biotechnological applications. Biol Res. 2013;46(4):395-405. http://dx.doi.org/10.4067/S0716-97602013000400011
- Kumar A, Singh S. Directed evolution: tailoring biocatalysts for industrial applications. Crit Rev Biotechnol. 2013;33(4):365-378. https://doi.org/10.3109/07388551.2012.716810
- Adrio JL, Demain AL. Microbial enzymes: tools for biotechnological processes. Biomolecules. 2014;4(1):117-139. https://doi.org/10.3390/biom4010117
- Song Z, Zhang Q, Wu W, Pu Z, Yu H. Rational design of enzyme activity and enantioselectivity. Front Bioeng Biotechnol. 2023;11: 1129149. https://doi.org/10.3389/fbioe.2023.1129149
- Ebert MC, Pelletier JN. Computational tools for enzyme improvement: why everyone can-and should-use them. Curr Opin Chem Biol. 2017;37:89-96.
 - https://doi.org/10.1016/j.cbpa.2017.01.021
- Tuan LQ. Rational protein design for enhancing thermal stability of industrial enzymes. HO CHI MINH CITY OPEN UNIVERSITY JOURNAL OF SCIENCE-ENGINEERING AND TECHNOLOGY. 2018; 8(1):3-17. https://doi.org/10.46223/HCMCOUJS.tech.en.8.1.340.2018
- 11. Bilal M, Iqbal HM, Guo S, Hu H, Wang W, Zhang X. State-of-the-art protein engineering approaches using biological macromolecules: A review from immobilization to implementation view point. Int J Biol Macromol. 2018;108:893-901. https://doi.org/10.1016/j.iibiomac.2017.10.182

https://doi.org/10.1016/j.ijbiomac.2017.10.182

- 12. Prasad S, Roy I. Converting enzymes into tools of industrial importance. Recent Pat Biotechno. 2018;12(1):33-56. https://doi.org/10.2174/1872208311666170612113303
- 13. Ismail AR, Kashtoh H, Baek KH. Temperature-resistant and solvent-tolerant lipases as industrial biocatalysts: Biotechnological approaches and applications. Int J Biol Macromol. 2021;187:127-142. https://doi.org/10.1016/j.ijbiomac.2021.07.101
- 14. Singh R, Langyan S, Rohtagi B, Darjee S, Khandelwal A, Shrivastava M, et al. Production of biofuels options by contribution of effective and suitable enzymes: Technological developments and challenges. Mater Sci Energy Technol. 2022;5:294-310. https://doi.org/10.1016/j.mset.2022.05.001
- Kumar A, Jangra A, Pramanik J. Application of Enzymes in Dairy Processing Industry: A Review. Curr Nutr Food Sci. 2022;18(5): 428-431. https://doi.org/10.2174/1573401318666220204093121
- 16. Ruan Y, Zhang R, Xu Y. Directed evolution of maltogenic amylase from Bacillus licheniformis R-53: Enhancing activity and thermostability improves bread quality and extends shelf life. Food Chem. 2022;381:132222. https://doi.org/10.1016/j.foodchem.2022.132222
- Sharma VK, Sharma M, Usmani Z, Pandey A, Singh BN, Tabatabaei M, et al. Tailored enzymes as next-generation food-packaging tools. Trends Biotechnol. 2022;40(8):1004-1017.
- https://doi.org/10.1016/j.tibtech.2022.01.009
- 18. Bhandari S, Poudel DK, Marahatha R, Dawadi S, Khadayat K, Phuyal S, et al. Microbial enzymes used in bioremediation. J Chem. 2021;2021(1):8849512. https://doi.org/10.1155/2021/8849512
- 19. Radley E, Davidson J, Foster J, Obexer R, Bell EL, Green AP. Engineering enzymes for environmental sustainability. Angew Chem Int Ed. 2023;62(52):e202309305. https://doi.org/10.1002/anie.202309305
- 20. Sharma A, Gupta G, Ahmad T, Mansoor S, Kaur B. Enzyme engineering: current trends and future perspectives. Food Rev Int. 2021; 37(2):121-154. https://doi.org/10.1080/87559129.2019.1695835
- 21. Tarafdar A, Sirohi R, Gaur VK, Kumar S, Sharma P, Varjani S, et al. Engineering interventions in enzyme production: Lab to industrial scale. Bioresour Technol. 2021;326:124771. https://doi.org/10.1016/j.biortech.2021.124771
- 22. Agarwal S, Sahu S. Safety and Regulatory Aspects of Food Enzymes: An Industrial Perspective.
- 23. Schmidt M, de Lorenzo V. Synthetic constructs in/for the environment: managing the interplay between natural and engineered Biology. FEBS Lett. 2012;586(15):2199-2206. https://doi.org/10.1016/j.febslet.2012.02.022
- 24. Singh N, Malik S, Gupta A, Srivastava KR. Revolutionizing enzyme engineering through artificial intelligence and machine learning. Emerg Top Life Sci. 2021;5(1):113-125. https://doi.org/10.1042/ETLS20200257
- Woodley JM. New frontiers in biocatalysis for sustainable synthesis. Curr Opin Green Sustain. 2020;21:22-26. https://doi.org/10.1016/j.cogsc.2019.08.006